

## HPLC Method Development for Simultaneous Estimation of Sodium Benzoate and Potassium Sorbate in Food Products

Sadaf K. Shaikh<sup>1</sup>, Dr. Mallinath S. Kalshetti<sup>2</sup> and Ravikant Y. Patil<sup>3</sup>

<sup>1,2,3</sup>Department of Quality Assurance, D. S. T. S Mandal's College of Pharmacy, Solapur, Maharashtra, India. Email: sdfshk94@gmail.com

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### ABSTRACT

A Reverse Phase-High Performance Liquid Chromatography method has been developed and validated for estimation of Sodium benzoate and Potassium sorbate in food products. The RP-HPLC method for Sodium benzoate and Potassium sorbate was developed using Luna C18 column (150 mm × 4.6 mm, 5 μm) as stationary phase and Acetonitrile: Sodium acetate buffer pH 4.3 (20:80) as mobile phase at 1.0ml/min flow rate and detection was carried out at 235nm and the methods were validated in accordance with ICH guidelines. Sodium benzoate and Potassium sorbate have linearity in the concentration range of 1-30μg/ml in with correlation coefficient ( $r_2= 0.999$  &  $r_2=0.999$ ) respectively. Sodium benzoate and Potassium sorbate eluted at 4.9 and 6.9 min respectively. The values of LOD were 0.59 and 0.39 and LOQ were 1.8 and 1.1 for sodium benzoate and potassium sorbate respectively. Results of assay and validation study are satisfactory. So, the methods can be successfully applied for the routine analysis of Sodium benzoate and Potassium sorbate.

Key Words: Sodium benzoate, Potassium sorbate, RP-HPLC Method, Food, Preservatives.

### 1. INTRODUCTION

In recent years, concern about the importance of food and diet quality has been growing, especially due to increase in the incidences of diseases that are directly or indirectly related to nutrition habits. As a natural outcome, analysis of food additives came into focus for the assessment of their harmful potentials and quantitative and qualitative value of risks related to their use.

A food additive is defined as a substance or a mixture of substances, which are generally added to processed foods for a specific purpose such as prevention of spoilage, conservation or fortification of color, flavor, texture or control of pH, moisture, crispness etc. Food additives may be divided into preservatives, artificial sweeteners, colorants, stabilizers etc.<sup>1</sup> Food and Agricultural Organization (FAO) and World Health Organization (WHO) have combined to form a Joint FAO-WHO expert Committee on Food Additives (JECFA) to evaluate and establish the acceptable daily intake (ADI) which is expressed in milligrams per kilograms of body weight per day. The most commonly used preservatives in foods are sodium benzoate and potassium sorbate, the use of which is limited by the regulating bodies. Sodium benzoate is a widely used food preservative, with an E number of E211. It is the sodium salt of benzoic acid. It is bacteriostatic and fungi static under acidic conditions. Commonly reported side effects of sodium benzoate include: eye irritations, urticaria, abdominal pain, asthma, hypertension, attention-deficit/hyperactivity disorder and vomiting. The ADI set by JECFA is 5 mg/kg bw.<sup>2</sup>

Potassium sorbate [potassium;(2E,4E)-hexa-2,4-dienoate] is the potassium salt of sorbic acid. It is primarily used as a food preservative and it is codified as E202. Potassium sorbate is used to inhibit growth of molds and yeasts in many foods, such as cheese, yogurt, soft drinks and fruit drinks, and baked goods. Allergic symptoms such as itching of the mouth, throat, eyes, skin as well as nasal congestion, runny nose and abdominal pain usually begin within 2 hours after coming in contact with the allergen. The ADI set by JECFA is 25 mg/kg bw.<sup>3</sup>

Thus, the analytical determination of these preservatives is not only important for quality assurance purposes but also for consumer interest and protection.

The present work deals with the development and characterization of a rapid and simple HPLC method for the simultaneous estimation of sodium benzoate and potassium sorbate in food products without an extensive sample pre-treatment. The proposed method has excellent performance characteristics, with a real potential to become a powerful tool for food safety.

## **2. EXPERIMENTAL**

### **2.1. MATERIALS AND METHODS**

Materials: Sodium Benzoate and Potassium Sorbate were obtained as a gift sample from Bimal Pharma Pvt. Ltd., Mumbai and GlaxoSmithKline Plc, Hyderabad respectively. Various food products were obtained from local market. HPLC grade Acetonitrile LiChrosolv®, Methanol LiChrosolv®, Water LiChrosolv®, Sodium acetate LiChrosolv® and Glacial acetic acid LiChrosolv® were purchased from Merck Specialities Pvt. Ltd. Mumbai.

#### **2.1.1. Instrumentation**

The analysis was performed using Younglin Acme 9000 series quaternary gradient pump SP930D. HPLC system with UV 730D UV-Visible detector and 20µl Rheodyne injector. The data was processed on Autochrom-3000 software. Column C18 (150mm×4.5mm,5µ) Phenomenex with UV method analysis was performed on UV-Visible Double Beam Spectrophotometer Shimadzu 1800.

All chemicals were weighed using Electronic Balance AY220 (Shimadzu, Japan). Mobile phase filtered through a Nylon 6,6 membrane 0.45 µm 47 mm filters (Pall India Pvt. Ltd., Mumbai) using vacuum pump. Ultra Sonicator (Microlean-103) was used for degassing the mobile phase. The solutions were filtered through 0.45µ syringe filter (Phenomenex).

#### **2.1.2. Chromatographic Conditions**

The chromatographic separation was performed using Analytical Column: Phenomenex C18 column (150 × 4.6 mm, 5 µm) using mobile phase Acetonitrile: Sodium acetate buffer Ph 4.3(20:80) at a flow rate of 1.0 ml/min with isocratic elution. The injection volume was 20 µl and the run time was 10 min. Detection was carried out at 235 nm.

#### **2.1.3. Preparation of Standard Stock Solution**

1. Standard Stock Solution of SB: 10mg of standard SB was weighed and transferred to a 10ml volumetric flask then dissolved in HPLC Water LiChrosolv® and the volume was made up to the mark with water to obtain conc. of 1000µg/ml of SB and labeled as 'Std Stock SB'.

2. Standard Stock Solution of PS: 10mg of standard PS was weighed and transferred to a 10ml volumetric flask then dissolved in HPLC Water LiChrosolv® and the volume was made up to the mark with water to obtain conc. of 1000µg/ml of PS and labeled as ‘Std Stock PS’.
3. Combined Standard Stock Solution of SB and PS: 1ml of ‘Std Stock SB’ and 1ml of ‘Std Stock PS’ was transferred to 10 ml volumetric flask and diluted to 10 ml with HPLC Water to get ‘Std Stock MIX SB PS’ (100µg/ml SB and 100µg/ml PS).

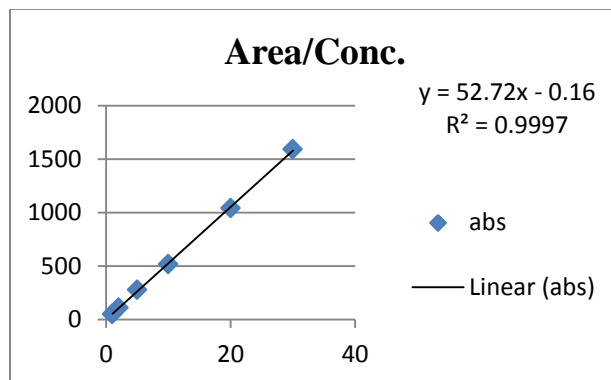


Fig. 1 Calibration curve of SB of RP-HPLC method

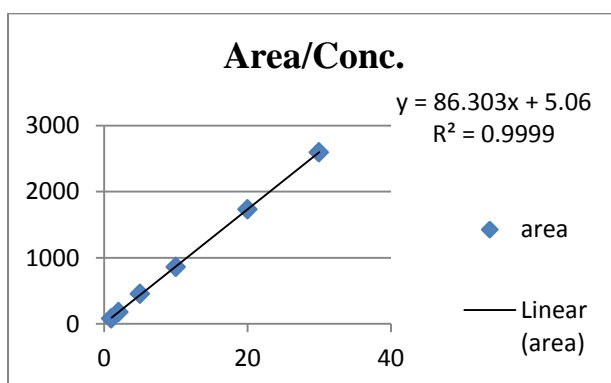


Fig.2 Calibration curve of PS of RP-HPLC method

## 2.2. Preparation of Samples

### 2.2.1. Sample A (Lauki-Amla Juice)

1ml of sample A was pipetted out and transferred to a 10ml volumetric flask. The volume was made up to the mark by HPLC water. Further dilution was done by pipetting out 1ml from the same volumetric flask and transfer to another 10ml volumetric flask. The volume was made upto the mark by HPLC water.

### 2.2.2. Sample B (Jam)

10gm of sample B was transferred to a 20ml beaker. The volume was made upto the mark by HPLC water. Further dilution was done by pipetting out 2ml from the same beaker and was transferred to a 10ml volumetric flask. The volume was made up to the mark by HPLC water.

### **2.2.3. Sample C (Cake)**

10gm of sample C was transferred to a 50ml beaker. The volume was made up to the mark by HPLC water. Further dilution was done by pipetting out 1ml from the same beaker and was transferred to a 10ml volumetric flask. The volume was made up to the mark by HPLC water.

### **2.2.4. Sample D (Fizz Juice)**

2ml of sample D was transferred to a 10ml volumetric flask. The volume was made up to the mark by HPLC water. All the samples were degassed in an ultrasonicator for 30 minutes and filtered through a nylon filter membrane (0.45 $\mu$ m) prior to use.

## **2.3. Preparation of Mobile Phase**

Mobile phase was prepared by mixing 20ml of Acetonitrile and 80ml of sodium acetate buffer (pH 4.3) and filtered through 0.45 $\mu$ m nylon filter using vacuum pump and ultrasonicated for 30 min for degassing.

## **2.4. Method Validation**

The developed analytical method as per the ICH Q2 (R1) guideline it is suitable for the intended purpose with respect to various parameters such as specificity, linearity, range, accuracy, precision, limit of detection, limit of quantification, robustness, system suitability.

### **2.4.1. Specificity**

The chromatogram of standard solution of mixture of SB and PS was compared with food products.

### **2.4.2. Linearity**

0.1, 0.2, 0.5, 1, 2 and 3ml of 'Std Stock MIX SBPS' were transferred in a series of 10ml volumetric flasks. The volume was made up to the mark with HPLC water to obtain the conc. of 1, 2, 5, 10, 20 and 30 $\mu$ g/ml of SB and 1, 2, 5, 10, 20 and 30 $\mu$ g/ml of PS. The solutions were filtered through 0.45 $\mu$  syringe filter and 20 $\mu$ l injected into the HPLC system and their chromatogram were recorded for 10mins under the chromatographic conditions as described above after getting a stable baseline.

Peak areas were recorded for all the peaks. Calibration curves of SB and PS were constructed by plotting the peak area of SB v/s conc. of SB and peak area of PS v/s conc. of PS, respectively. The correlation coefficient ( $r^2$ ) of least square linear regression for SB and PS was calculated.

### **2.4.3. Range**

The range of analytical method was decided from the interval between upper and lower level of calibration curves.

#### **2.4.4. Precision**

10 $\mu$ g/ml of SB and 10 $\mu$ g/ml of PS solution was filtered through 0.45 $\mu$  syringe filter and 20 $\mu$ l injected into the HPLC system and its chromatogram was recorded under the same chromatographic conditions after getting a stable baseline. Peak area was recorded. The procedure was repeated for six times.

#### **2.4.5. Accuracy**

1, 0.5, 0.5, and 2ml from sample A, B, C, and D respectively was taken and transferred to 10ml volumetric flasks each. The volume was made up to the mark with mobile phase. Similarly, another 4 sets were prepared with the addition of 1ml of 'Std Stock MIX SBPS' in each. All the solutions were filtered through syringe filter and injected into the HPLC system and their chromatograms were recorded under the same chromatographic conditions after getting a stable baseline. Peak areas were recorded and percent recoveries were calculated.

#### **2.4.6. Limit of Detection**

LOD calculated by the following formulae.

$LOD = 3.3(SD/S)$  Where, SD- Standard deviation; S- Slope of Curve

#### **2.4.7. Limit of Quantification**

LOQ calculated by the following formulae.

$LOQ = 10(SD/S)$  Where, SD- Standard deviation; S- Slope of Curve.

#### **2.4.8. Robustness**

Combined standard solution of SB (10 $\mu$ g/ml) and PS (10 $\mu$ g/ml) was prepared and analyzed at different flow rates (0.9, 1.0, 1.1 ml/min) and different wavelengths (234, 235, 236nm) separately.

#### **2.4.9. System Suitability**

Chromatograms were studied for different parameters such as tailing factor, resolution and theoretical plates to see that whether they comply with the recommended limit or not.

### **3. RESULTS AND DISCUSSIONS**

In order to develop RP-HPLC method for combination of Sodium Benzoate and Potassium sorbate, the chromatographic conditions were optimized in order to find the best conditions. Different mobile phase like acetonitrile, methanol, water and buffer in varying proportions of mobile phases were tried for better resolution. After several combinations of mobile solvents with stationary phase C18, the above method has been optimized i.e. acetonitrile: sodium acetate buffer (Ph 4.3) in the ratio of 20:80 respectively using C18 column which has given good resolution, capacity factor, and acceptable system suitability. Chromatographic peak of both drugs are identified by overlaying individual drug with chromatograph of mixture. Both drugs eluted within 10mins which reduces the analysis time and cost.

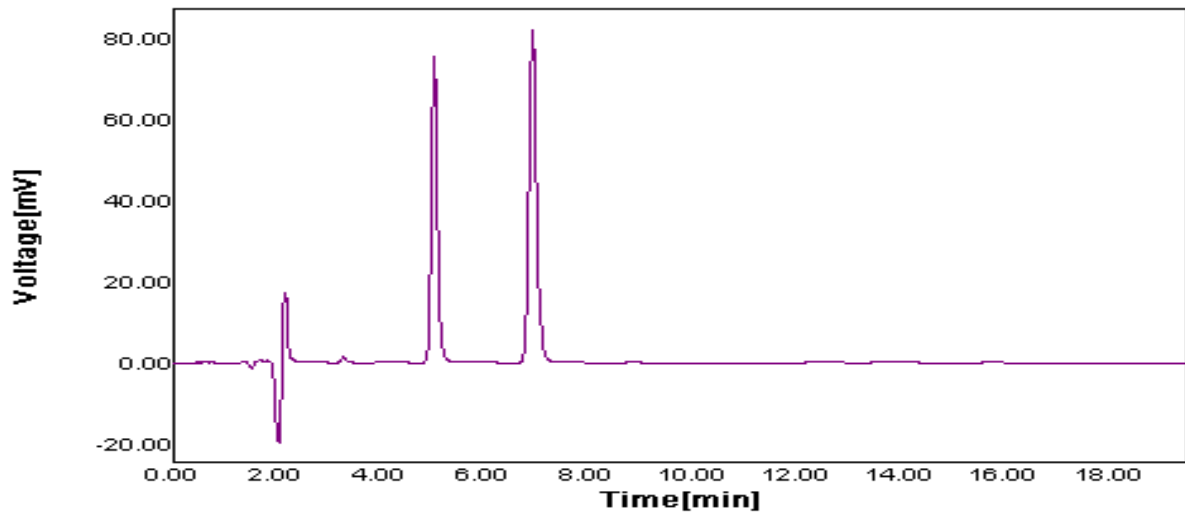


Fig.3 Chromatogram of combination of SB (10µg/ml) & PS (10µg/ml) in optimized chromatographic conditions

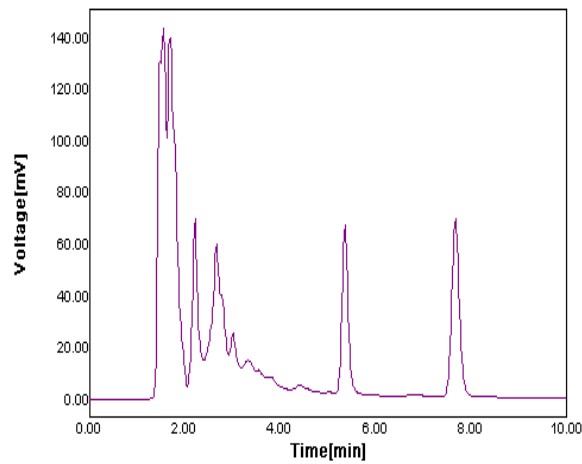


Fig.4 Chromatogram of Juice sample

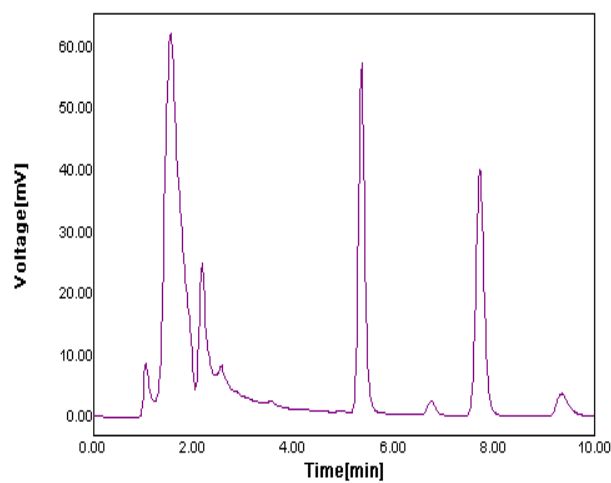


Fig. 5 Chromatogram of Jam sample

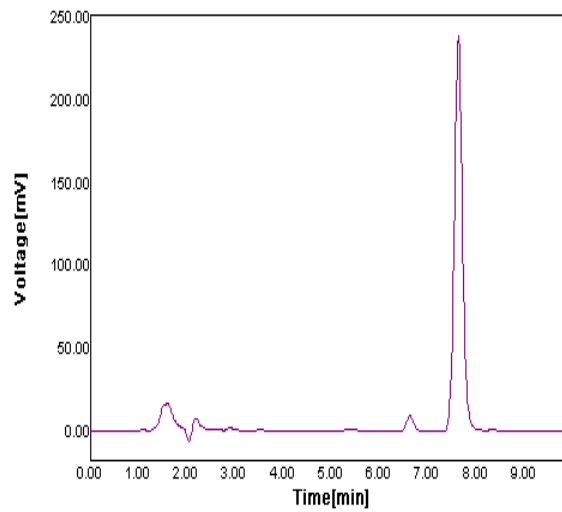


Fig. 6 Chromatogram of Cake sample

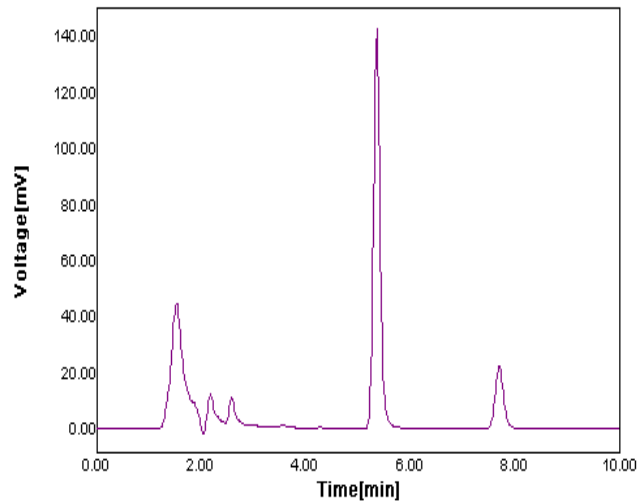


Fig. 7 Chromatogram of Fizz juice sample

### 3.1. ACCURACY

Table 1- Accuracy for RP-HPLC Method

Sr. No.	Sample No.	Level of Recovery	Amount of Sample (ml)	Amount of Standard Drug Added (µg/ml)		Total Amount Found (µg/ml)		Amount Recovered (µg/ml)		% Recovery	
				SB	PS	SB	PS	SB	PS	SB	PS
1	A	0%	1	10	10	11.38	8.38	0	0	0	0
		100%				11.38	8.38	19.86	17.35	97.7	98.8
2	B	0%	0.5	10	10	2.31	1.15	0	0	0	0
		100%				2.31	1.15	11.19	10.40	101.1	102.2

3	C	0%	0.5	10	10	-	14.21	0	0	0	0
		100%				-	14.21	8.61	24.39		98.6
4	D	0%	2	10	10	23.86	2.68	0	0	0	0
		100%				23.86	2.68	32.2	11.91	98	102.2

### 3.2. PRECISION

Table 2- Results of Repeatability Study for SB and PS

Inj.	Peak Area(mV)of SB	Peak Area(mV)of PS
1	516	860
2	500	838
3	524	846
4	520	860
5	525	860
6	524	865
SD	9.5	10.3
RSD	1.83	1.2

### 3.3. LIMIT OF DETECTION

Table 3 - Limit of Detection data of SB and PS

	SB	PS
LOD (µg/ml)	0.59	0.39

### 3.4. LIMIT OF QUANTITATION

Table 4 - Limit of Detection data of SB and PS

	SB	PS
LOQ (µg/ml)	1.80	1.19



### 3.5. ROBUSTNESS

Table 5- Result of Robustness Study: Variation in Flow Rate (ml/min)

Flow Rate (ml/min)	Analyte	Retention Time (min)	Tailing Factor (T)	Theoretical Plates (N)	Resolution (R)
0.9	SB	5.2	1.2	8457	2.9
	PS	7.5	1.1	10987	8.7
1.0	SB	4.8	0.8	8823	2.6
	PS	6.8	1.1	10713	8.6
1.1	SB	4.4	1.2	8400	1.3
	PS	6.3	1.1	10912	8.9

Table 6 - Result of Robustness Study: Variation in Wavelength (nm)

Wavelength (nm)	Analyte	Retention Time (min)	Tailing Factor (T)	Theoretical Plates (N)	Resolution (R)
234	SB	4.8	1.3	7428	065
	PS	6.9	1.1	10624	8.59
235	SB	4.8	0.8	8823	2.69
	PS	6.9	1.1	10713	8.6
236	SB	4.8	1.1	9162	1.3
	PS	6.9	1.1	11197	9

### 3.6. SYSTEM SUITABILITY TESTING

Table.7 Results of System Suitability Parameters

Analyte	Retention Time (min)	Tailing Factor (T)	Theoretical Plates (N)	Resolution (R)
SB	4.8	0.84	8823.5	2.69
PS	6.9	1.18	10713.2	8.60
Required limits	--	T < 2	N > 2000	R > 2

#### 4. CONCLUSION

This research was aimed to extract and evaluate the amount of preservatives in various food products. The proposed method was RP-HPLC which was found to be appropriate due to its simplicity, reliability, sensitivity, rapidness, and selectivity for detection at very low concentrations. It could recognize the preservatives at one wavelength in less than 10 mins and involves minimal sample preparation. The recoveries were within 97-102 % which implies that the method is accurate. It was found that Sample C (cake) exceeded the permissible limit of preservative and was not in accordance with the limits set by Food Safety and Standards Authority of India and CODEX STAN 192-1995. However the other 3 samples complied with the limits satisfactorily.

In conclusion, the HPLC method is simple, accurate, reliable and reproducible method for simultaneous estimation of SB and PS in food products. Optimum values of the retention time, pH, flow rate and mobile phase ratio for good separation of the analytes were determined. The optimized HPLC methods were validated and based on the results obtained the proposed method is found to be linear, precise and accurate. Validation data demonstrates that this method can be used in routine analysis of sodium benzoate and potassium sorbate in various food products.

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